considered outer sphere in nature. Davies²² has analyzed the variations in rate constants for the latter reactions in terms of outer-sphere precursor formation and the attainment of a species in which the distribution of OH⁻ in the inner sphere of cobalt(II1) is optimal for hydroxide-mediated electron transfer within the precursor complex. **A** specific interaction of this type is consistent with the present system.

The slower reaction $(k_3K_h = 935 \text{ s}^{-1})$ for the oxidation of Ni(cyclam) compared with other anionic or neutral substrates (e.g., $I^-, k_3K_h = 2.9 \times 10^3 \text{ s}^{-1}$) may reflect either the higher redox potential of the Ni(II)L/Ni(III)L couple (0.95 V) or the fact that electrostatic repulsion terms will be greater in the case of cationic reductants. The former argument does not explain the relatively low reaction rates for the Co^{2+} , Mn²⁺,

(22) Rickman, R. A.; Sorensen, R. L.; Watkins, K. 0.; Davies, G. *Inorg. Chem.* **1977,** 16, 1570. (23) Davies, **G.** *Inorg. Chem.* **1971,** *10,* 1157.

and $Fe²⁺$ reactions.²³ Differences in the rates will be however expected if a specifically oriented precursor is formed prior to the electron transfer. **A** series of preequilibria22 describing such a process have been suggested as contributing to the compensating variation in $\Delta \vec{H}^*$ and ΔS^* observed in these reactions. The fact that a metal complex ion is involved rather than the smaller aquo ions will have a bearing on these effects.

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Registry No. [Ni"L]*+, 46365-93-9; Co(III)(aq), 15275-05-5; $[NiL(OH₂)₂]$ ³⁺, 72360-42-0; $[NiL(MeCN)₂]$ ³⁺, 47099-85-4; $COOH²⁺(aq)$, 68842-90-0.

Contribution from the Istituto di Chimica Generale ed Inorganica, Facolta di Farmacia, University of Florence, 50121 Florence, Italy, and Laboratorio per lo Studio dei Composti di Coordinazione del CNR, Florence, Italy

pH-Dependent Properties of a CoN4(0H2) Chromophore: A Spectroscopic Model of Cobalt Carbonic Anhydrase

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The complexes $[Co(H₂O)L] (ClO₄)₂$ and $[Co(N₃)L] ClO₄$, where L stands for tris[(3,5-dimethyl-1-pyrazolyl)methyl]amine, have been prepared and characterized. The aqua complex has been found to have two acidic groups in the pH range 5.5-9.5, which have been attributed to the bound water ($pK_a \approx 9$) and to the apical nitrogen ($pK_a \approx 7.4$). The behavior of this compound as well as of its azide derivative has been compared with that of cobalt(I1)-substituted carbonic anhydrase; in particular, it is shown that the deprotonation of the coordinated water molecule can by itself cause a dramatic change in the electronic spectra of the cobalt(I1) chromophore and a strong reduction of the anion affinity for the solvent coordination site, both puzzling properties of the above enzyme.

Introduction

Carbonic anhydrase is a zinc(I1) metalloenzyme with a water molecule in the metal coordination sphere, whose catalytic activity depends on an acid-base equilibrium with pK_s of about 7.2 The same equilibrium also controls the inhibiting ability of anions like NCO⁻, N₃⁻, halides, etc.² The identity of the acidic group involved is still controversial:³ among various possibilities, the coordinated water molecule has been often regarded as a reasonable candidate.⁴

Cobalt(I1) can be successfully substituted to the native metal ion yielding a still active enzyme with similar acid-base properties.⁵ The electronic spectra of the latter derivative are strongly pH dependent;^{2a} although they are probably sensitive to more than one acid-base equilibrium⁶ (actually about four or five deprotonations take place in the enzyme in the pH range $5-9^{2b}$), the major variation can again be ascribed to a single acidic group with pK_a of about 7. However, the observed spectral changes cannot be taken as diagnostic of ionization of the coordinated water molecule. Indeed, none of the few known cobalt(I1) complexes with a single water molecule bound in water solution⁷⁻¹¹ are suitable to reproduce the spectral features and variations observed in the cobalt enzyme.

In order to investigate the acid-base properties of the coordinated water molecule, the influence of the water deprotonation on the electronic spectra, and the possibility of substitution of the H_2O or OH groups by other ligands, we have

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prepared and characterized the complexes $[Co(H₂O)L](ClO₄)₂$ and $[Co(N₃)L]ClO₄$, where L is tris $[(3,5\textrm{-dimethyl-1-}$ pyrazolyl)methyl] amine.

This ligand has been chosen for the following reasons: (i) pyrazolyl-containing ligands are known to stabilize low oxidation states¹² and therefore should protect cobalt(II) from

- (1) (a) Istituto di Chimica Generale ed Inorganica, Facolts di Farmacia, University of Florence, and Laboratorio per lo Studio dei Composti di Coordinazione del CNR. (b) Istituto di Chimica Generale *ed* Inorganica, University of Florence.
- (2) (a) Lindskog, **S.;** Henderson, L. E.; Kannan, K. K.; Lijas, **A,;** Nyman, P. 0.; Strandberg, B. *Enzymes, 3rd Ed.* 1970-1976 **1971,** *5.* **(b)** Coleman, J. E. "Inorganic Biochemistry"; Eichhorn, *G.* I., Ed.; Elsevier: London, 1973; Vol. 1.
- (3) Pocker, *Y.;* Sarkanen, *S. Adv. Enzymol. Relar. Areas Mol. Biol.* **1978,** 47, 149.
- (4) Lindskog, *S.;* Coleman, J. E. *Proc. Narl. Acad. Sci. U.S.A.* **1973** *70,* 2505.
- (5) Thorslund, **A,;** Lindskog, *S. Eur.* J. *Biochem.* **1967,** *3,* 117.
- (6) Bertini, I.; Luchinat, *C.;* Scozzafava, A. *Inorg. Chim. Acra* **1980,** 46,
-
- 85. (7) Wolley, **P.** *Nature (London)* **1975,** *258,* 677. (8) Meier, P.; Merbach, **A.;** Biirki, **S.;** Kaden, T. **A.** *J. Chem. SOC., Chem. Commun.* **1977,** 36.
- **(9)** Paoletti, P.; Ciampolini, M.; Sacconi, L. *J. Chem. Soc.* **1963,** 3589.
- **(IO)** Dei, A.; Paoletti, P.; Vacca, **A.** *Inorg. Chem.* **1968, 7,** 865. (11) Vacca, A.; Paoletti, P. *J. Chem. SOC. A* **1968.** 2378.
-
- (12) Trofimenko, *S. Chem. Rev.* **1972,** 72,498.

^{*a*} Reference values of 2:1 electrolytes in 10⁻³ mol dm⁻³ CH₃OH solutions give an average $\Lambda_M = 190$ S cm² mol⁻¹.¹⁸ Conductivity of ethylenediamine dihydrochloride in 10^{-3} mol dm⁻³ water solution is 240 S cm² mol⁻¹.

oxidation even at high pH values; (ii) the tripodal geometry **of** the ligand and the three methyl groups in the 3-positions should favor five-coordination with a single water molecule; (iii) pyrazoles bear some similarity to imidazoles, which are the coordinating groups from the protein in carbonic anhydrase.^{2a}

Experimental Section

The ligand tris[(3,5-dimethyl- 1-pyrazolyl)methyl]amine was synthesized by allowing $N(CH_2Cl)_3$ and potassium 3,5-dimethylpyrazolate to react as already reported.¹³ $[Co(H₂O)L](ClO₄)₂$ was obtained upon evaporation of an ethanol solution containing $Co(H_2O)_6(CIO_4)_2$ and the ligand in 10% excess. $[Co(N_3)L]ClO_4$ was prepared by adding an equimolar amount of potassium azide to a methanol solution of the aqua complex.

The complexes were recrystallized from ethanol or methanol, filtered **off, and** washed with petroleum ether. The analytical data are reported in Table I. The presence of water in the $[Co(H₂O)L](ClO₄)_{2}$ complex is confirmed by infrared spectra which show bands in the region 3200-3400 cm⁻¹ (OH stretching) and 1600-1700 cm⁻¹ (HOH bending).

The conductivity data were obtained with a WTW Model LBR/B conductance bridge. Concentrations of the solutions were approximately 10⁻³ mol dm⁻³. Magnetic measurements were performed with the Faraday method by using the apparatus and the techniques already reported." Infrared spectra were obtained with a Perkin-Elmer Model 283 grating spectrophotometer on samples prepared as Nujol mulls.

Potentiometric measurements were performed with a potentiometer, **Orion Model 801, in the pH** range $9.2-5.0$, with 10^{-3} mol dm⁻³ complex solutions.

The electronic spectra were recorded on a Cary 17D spectrophotometer. The spectra in the near-infrared region were recorded in D_2O or deuteriomethanol solutions. The binding of N_2 to the cobalt complex in the pH range 6-9.5 was followed from the spectral variations in the visible region.

The 80-MHz ¹H NMR spectra of the complexes in D_2O or CD_3OD were recorded on a Varian CFT 20 spectrometer operating at 20 °C. Pulses of 90° were employed with repetition times of 0.05-0.1 s. The short relaxation times of the protons of paramagnetic complexes permitted enhancement of the ligand signals relative to the signals of the solvent protons, which are almost saturated under these conditions. Chemical shifts were measured from external tetramethylsilane. T_1 measurements were performed by using the inversion recovery method; water proton relaxation measurements were performed as previously described.¹⁵ The reported pH values in D_2O solutions are uncorrected, and they correspond to $pD -0.4$.¹⁶

The aqua complex was tested as catalyzer of the CO₂ hydration reaction following the method of Roughton and Booth." No increase in the hydration rate is observed up to 10^{-3} mol dm⁻³ complex concentration at pH 8.8.

Results and Discussion

The complex $[Co(H₂O)L](ClO₄)₂$ is high-spin: its reflectance spectrum shows absorption at 6.8, 13.6, and **19.2** cm-' \times 10⁻³ (Figure 1). The analogies with other $[CoXL]$ ⁺ complexes $(X =$ halides, pseudohalides) previously reported¹³ lead us to assign the complex a five-coordinate structure. The

(15) Bertini, I.; Canti, G.; Luchinat, C.; Scozzafava, **A.** *J. Am. Chem. Soc.* **1978,** *100,* 4873.

Figure 1. Electronic spectra of the $[Co(H, O)L](ClO₄)$, complex in water solution at pH 7.5 **(A)** and 9.6 (B) and as a solid (C) and electronic spectrum of solid $[Co(N_3)L]ClO_4$ (D).

Figure 2. pH dependence of the electronic spectra of the $[Co(H₂-)]$ $O(L(C1O_4)_2)$ complex in water: from the top, pH 9.55, 9.3, 9.15, 8.9, 8.7, 8.3, 7.9, 7.7, 7.6, 7.2, 6.2, 5.0. The pH of the solid line (8.3) is that resulting from dissolving the complex in water at 10^{-3} mol dm⁻³ concentration. The insert shows the variation of ϵ_{490} with pH.

complex is soluble in water, in methanol, and in most polar organic solvents. Conductivity measurements in water and methanol provided values indicative of 2:1 electrolyte¹⁸ (Table

⁽¹³⁾ Mani, F.; Scapacci, *G. Znorg. Chim. Acfa* **1980,** *38,* 151.

⁽¹⁴⁾ Sacconi, L.; Bertini, I.; Mani, F. *Znorg. Chem.* **1968, 7,** 1417.

Figure 3. Electronic spectra of the isoenzyme B of cobalt(I1) human carbonic anhydrase at pH 6.1 **(A), 7.8** (B), and 9.5 (C).

I), in agreement with the proposed structure. In water solution at about pH **7.5** and in methanol solution the electronic spectra are very similar to that recorded on the solid sample (Figure **1).** The spectrum obtained in water solution is dramatically pH dependent, as shown in Figure **2.** The analysis of the spectral data indicates the presence of two acid-base equilibria (Figure **2,** insert): upon acidification of the aqua complex (full-line spectrum) the color of the solution fades, indicating that the apical nitrogen is protonated and six-coordinated polyaqua complexes are formed;19 upon increase of pH the spectrum of a new species appears, indicating the presence of a further single acid-base equilibrium, as shown by the presence of two isosbestic points, whose pK_a is estimated to be 8.9 ± 0.2 .²⁰ This equilibrium should be ascribed to the deprotonation of the bound water molecule to yield a [Co- $(OH)L$ ⁺ chromophore; the electronic spectrum of the hydroxo complex (Figure l), although markedly different from that of the conjugated acid, is still consistent with five-coordination.

The electronic spectra of both the low- and the high-pH forms of cobalt carbonic anhydrase are reported in Figure 3: a striking similarity is apparent with the spectra of the present $[Co(H₂O)L]²⁺$ and $[Co(OH)L]⁺$ chromophores. The coordination geometry of both forms of carbonic anhydrase is generally believed to be pseudotetrahedral, with a $N₃O$ donor set;^{1a} the presence of a fifth nitrogen donor in the case of the model complex results in an increase in energy of the highest F-F transition and in a decrease of the overall molar absorbance. Nevertheless, the deprotonation of the water molecule in the model complex successfully mimics the spectral variations of the cobalt enzyme with pH, making less speculative the occurrence of the same process in the enzyme.

Of course this finding does not allow any further inference on the catalytic mechanism besides providing some support to those mechanisms which involve H_2O and OH^- along the process.⁴

Another property of the enzyme to be accounted for is the decrease in affinity of anions for the metal site with increasing

(18) Geary, W. *Coord. Chem. Reu.* 1971, **7, 81.**

Figure 4. Electronic spectra of 10^{-3} mol dm⁻³ $[Co(H_2O)L] (ClO_4)_2$ with increasing amounts of NaN₃: from the bottom, 0, 1.5 \times 10⁻³, 3.0×10^{-3} , 9.0×10^{-3} , 0.2 mol dm⁻³. The insert shows the pH dependence of the apparent affinity constant of N_3 ⁻ for the [Co- $(H₂O)L]²⁺ complex.$

pH. When N_3^- is added to water solutions of the [Co- $(H₂O)L]²⁺ complex, a new spectrum is obtained similar to that$ of the $[Co(N_3)L]ClO_4$ complex (Figure 1), which has been isolated in the solid state (see Experimental Section). When the spectra are recorded at increasing amounts of N_1 ⁻ (Figure **4),** two isosbestic points appear, and the affinity constant for the equilibrium

$$
[C_0(H_2O)L]^{2+} + N_3^- \rightleftharpoons [C_0(N_3)L]^{+} + H_2O
$$

could be determined through a computer treatment of the spectrophotometric data. The apparent affinity constant K_{apo} at pH $\dot{8}$.1 is 280 ± 20 dm³ mol⁻¹. Such analysis indicates also that only **1:l** adducts are formed. Although the spectrum of the N_3 ⁻ adduct is not pH dependent, the affinity constants are, as shown in Figure **4** (insert). The dotted line is calculated from the relationship

$$
K_{\rm app} = \frac{K}{\left(1 + \frac{[H^+]}{K_{\rm al}}\right)\left(1 + \frac{K_{\rm a2}}{[H^+]}\right)}
$$

with the assumption of $K = 420$, negligible affinity for the high pH form, $pK_{a1} = 7.5$, and $pK_{a2} = 8.8$.

The above data indicate that N_3 reacts only with the acidic form, although the possibility of a small affinity for the high-pH form cannot be ruled out. Again, the data confirm that competition between anions and hydroxide ions can be a reasonable hypothesis also in the enzyme. Despite the many similarities between the model complex and the cobalt enzyme from the chemical point of view, the former derivative does not show any catalytic activity with respect to $CO₂$ hydration (see Experimental Section). Obviously, the catalytic efficiency of an enzyme depends on the presence of many concomitant structural requirements, of which the metal chromophore is only a component.

The ¹H NMR spectra of the $[Co(H₂O)L](ClO₄)₂$ complex in **D20** at pH **7.8** and **9.6** and the spectrum obtained in

The protonation rate is very low (about 1 h to reach constant pH), and the pK_a is estimated to be 7.4 \pm 0.2. Tertiary amine nitrogens have pK_a **values of around IO, which are lowered** upon **coordination to a metal ion, depending** on **the stability of the complex. On the other hand, the pyrazole nitrogens are expected to have pK, values** of **around 2.**

⁽²⁰⁾ The uncertainty of the latter value is due to precipitation of hydroxo species at high pH values: above pH 9.6 the amount of complex in solution becomes too low to perform any measurement. Potentiometric titrations yielded $pK_a = 9.1 \pm 0.1$ **for the above equilibrium; the whole titration in the pH range 9.2-5.0 required 1.7 mol** of **protons/mol** of **complex.**

Figure 5. ¹H NMR spectra of $[Co(H₂O)_L](ClO₄)$, in $D₂O$ at the uncorrected pH values of **9.6** (A) and **7.8** (B) (for the determination of pH **see** the Experimental Section) and in the presence of IO-' mol $dm^{-3} N_3$ ⁻ (C).

presence of 10^{-1} mol dm⁻³ NaN₃ are reported in Figure 5. The latter spectrum consists of four signals of intensities 2:1:3:3, which are assigned to the $CH₂$ group, to the C4 proton, and to the 5-CH₃ and the 3-CH₃ groups, respectively.²¹ In the former spectra three signals of relative intensities 1:2:3 are observed; the 3 -CH₃ signal is apparently underlying the intense peak of HDO protons. The large isotropic shift differences observed in the signals of the aqua complex at the two pH values, particularly evident in the CH₂ protons, point out that a considerable conformational change accompanies the deprotonation of the metal-bound water molecule. The **NMR** spectrum at high pH is somewhat similar to that of the azide derivative. The occurrence of some stereochemical rearrangement upon water deprotonation is also consistent with the variation of the electronic spectra, since switching from H₂O to OH⁻ is not expected to cause any major change in the energy levels, the two groups being close in both the spectrochemical and nephelauxetic series.²²

Longitudinal relaxation rates of the **'H** signal of water in cobalt enzyme solutions have been found to be pH independent.¹⁵ Actually, a decrease would be expected on passing from $H₂O$ to $OH⁻$ owing to the decrease in the number of exchangeable protons interacting with the paramagnetic center, unless the metal-proton distance sensibly decreases in the latter case. In the present aqua complex, the T_1 values of water protons decrease from 3.5 to 2.0 s when the paramagnetic complex is dissolved in water at 1.0×10^{-3} mol dm⁻³ concentration. The measured effect is indeed substantially pH independent, although too small to be safely compared with the enzyme results.

76773-06-3; isoenzyme B, **9001-03-0;** Co, **7440-48-4. Registry No.** $[Co(H_2O)L](ClO_4)_2$, 76773-04-1; $[Co(N_3)L](ClO_4)$,

(22) Jerrgensen, C. K. "Modern Aspects of Ligand Field Theory"; North-Holland Publishing Co.: Amsterdam, 1971.

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Electrochemical Study of a Trinuclear Copper (II) System: The Cu^{II}₂Cu^{III}–Cu^{II}₃ Couple and Its Inhibition by Proton

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An electrochemical study of a group of spin-doublet $(S = \frac{1}{2})$ trinuclear copper(II) complexes, $[Cu₃O_p(OH)_{1-p}(lig$ and)₃](ClO₄)_{2-p} [0 $\leq p \leq 1$; ligand = RNC(R')C(R')NO⁻; R = Et, n-Pr, n-Bu, Ph; R' = Me, Ph], is reported. In general, the complexes as isolated contain both Cu₃O and Cu₃OH cores held by three peripheral oximato bridges of the ligand. Cyclic voltammetry (both dc and ac) and constant potential coulometry establish that only the Cu₃O core displays the novel one-electron-transfer process Cu^{III}Cu^{II}₂ + e⁻ \rightleftharpoons Cu^{II}₃ with E^o₂₉₈' in the at a platinum working electrode. The rare Cu(II1)-Cu(I1) mixed-valence species thus discovered have an intense electronic absorption band at ~830 nm. The heterogeneous rate constants of electron transfer are reported. Base (NEt₃) converts Cu₃OH to a Cu₃O species in an equilibrium deprotonation reaction. On the other hand an acid (HClO₄) brings about absorption band at ~830 nm. The heterogeneous rate constants of electron transfer are reported. Base (NEt₃) converts Cu₃OH to a Cu₃O species in an equilibrium deprotonation reaction. On the other hand an acid (HClO and proton transfer is noted.

The function of copper oxidases and oxygenases is dependent on the variable valency of the metal ion.^{$I₋₃$} This has stimulated much current interest in electron-transfer studies of C_{ij}^i and C_{ij}^i and C_{ij}^i and C_{ij}^i are C_{ij}^i and C_{ij}^i are C_{ij}^i and C_{ij}^i are C_{ij}^i are C_{ij}^i are C_{ij}^i are C_{ij}^i are C_{ij} **Cum at a common interval in such studies.** The electron-transfer steps C^1 C^2 C^1 C^1 C^2

- **(3) Pecht, I.; Ferver, 0.; Goldberg, M.** *Adu. Chem. Ser.* **1977,** *No.* **262, 179.**
- **(4) Patterson, G. H.; Holm, R. H.** *Bioinorg. Chem.* **1975,** *4,* **257.**

as schematically summarized in **1** and **2.** Though unknown

at present, given proper ligand environment, a similar triangle

⁽²¹⁾ Horrocks, W. DeW. Jr. "NMR of Paramagnetic Molecules"; La Mar, G. N., Horrocks, W. DeW., Jr., Holm, R. H., Eds.; Academic Press: New York, 1973. Bertini, I. J. Mol. Struct. 1978, 45, 173.

⁽¹⁾ Fee, J. A. *Sfrucf. Bonding (Berlin)* **1975,** *23,* 1.

⁽²⁾ Malmstrom, B. *G.;* **Andreasson, L. E.; Reinhammer, B.** *Enzymer, 3rd ed.* **1975,** *12,* **507.**

⁽⁵⁾ Dockal, E. **R.; Jones, T. E.; Sokol, W. F.; Engerer, R. J.; Rorabacher, D. B.; Ochryrnoweycz, L. A.** *J. Am. Chem. SOC.* **1976,** *98,* **4322.**